

Cryptosporidium Methods - Best Practices and Useful Tips

Teaching Grandma to Suck Eggs...

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Overview

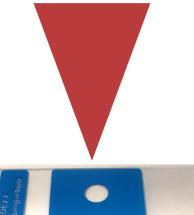
- Method Overview
- Sample Filtration
- Filter Elution
- Eluate Concentration
- Sample Purification
- Sample Staining
- A Word about Controls
- The Key to Consistency



Method Overview

- Sample Collection
 - filtration (10L raw 1000⁺L finished)
- Filter Elution
 - volume reduced to 250-1200ml
- Eluate Concentration
 - volume reduced to ~10ml
- Target Isolation (Purification)
 - volume reduced to ~50-100μl
- Target Identification
 - microscopy (9-15mm well slide)







Sample Filtration







Sample Filtration

- Be consistent
 - volume, flow rate and pressure
- Keep equipment in good order
 - reusable or disposable?
- Check tubing for wear and tear (replace if required)
- Mix the sample continuously
- Rinse the sample container
 - 2 x 500ml is better than 1 x 1L (2 x 1L is even better)
 - DO NOT use rinses containing detergents







Your choice of sampling tubing can dramatically affect the performance of the method!

For example, (oo)cysts stick to peroxide cured but not platinum cured silicone tubing!



Filter Elution







Filter Elution

- Be consistent
 - automate where possible
 - use consistent methodology
- Buffer solutions
 - make them consistently
 - use pre-prepared reagents
 - make them regularly
 - use clean equipment (manufacture & storage)
 - keep them out of direct sunlight
- Maintain equipment appropriately



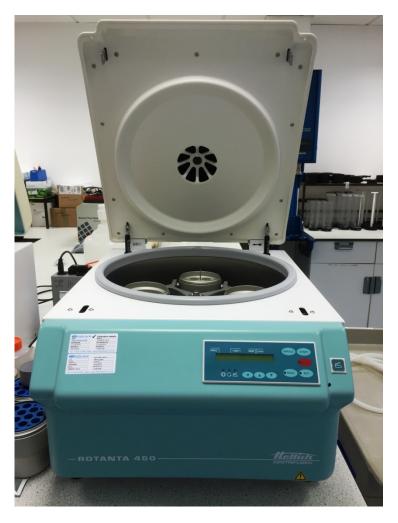




Buffer solutions can easily become contaminated with bacteria which can use surfactants as a carbon source!



Eluate Concentration







Eluate Concentration - Centrifugation

- Be consistent
- Location, location
 - solid ground and floor standing are best
- Maintenance
 - servicing but also general maintenance
- Balancing
 - balance tubes, supports & buckets & run a full rotor
 - don't add liquid to the buckets
- Speed & time are critical (rpm ≠ rcf)
- Temperature affects centrifugation efficiency
 - for multiple runs consider refrigeration or run two units
- Don't leave samples sitting after centrifugation







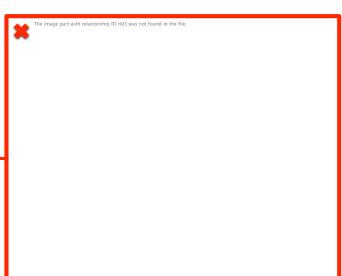
The zero brake setting on some centrifuges doesn't mean no braking!

With some models, the brake will engage initially and then cut out at a certain speed!



Eluate Concentration - Aspiration







Eluate Concentration - Aspiration

- Be consistent
 - maintain a consistent vacuum strength
 - limit aspiration rate
 - ▶ 200ml/min
- Keep it stable
 - don't move too far
- Use wide bore pipette tips
- Keep the pipette at the surface of the liquid
- Keep your eye on the ball
- Don't rush it





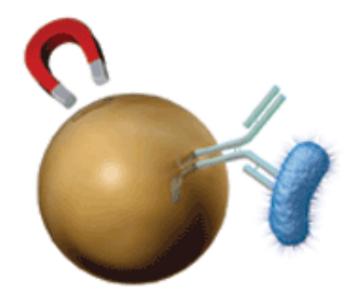


The use of wide-bore pipette tips can dramatically reduce the shear force during aspiration reducing the risk of pellet loss!



Sample Purification - IMS







Sample Purification - IMS

- Be Consistent
 - incubation time
 - rotation speed
 - rotation angle (it does matter)
- Leighton tubes
 - the flat side should be flat
 - don't use tubes with a short window
 - don't use them if they are cracked or crazed
 - replace caps regularly (make them disposable)





Sample Purification – IMS cont...

- During transfer from the Leighton tube
 - use plastic Pasteur pipettes (not glass) for transfers
 - 3 transfer rinses are better than 2
 - be careful with the final volume
 - use low-adhesion micro-centrifuge tubes



- During magnetic separation
 - use the recommended magnets
 - ensure the magnets are in good condition
 - ensure the magnetic strip is in the correct position
 - keep things moving
 - aspirate quickly and consistently



Sample Purification – IMS cont...

- During acid dissociation
 - use standard solutions of acid and alkali
 - replace the solutions often
 - use reverse pipetting for small volumes
 - take care vortexing tubes (keep the beads at the base)



- Avoid a pH imbalance during double acid dissociation
 - there are several options
 - apply 5µl alkali to 50µl acid twice
 - apply the 10µl of alkali to 100µl of acid at the end
 - use two slides





When transferring a sample from a Leighton tube, a light coloured background can help you visualise the remaining beads!





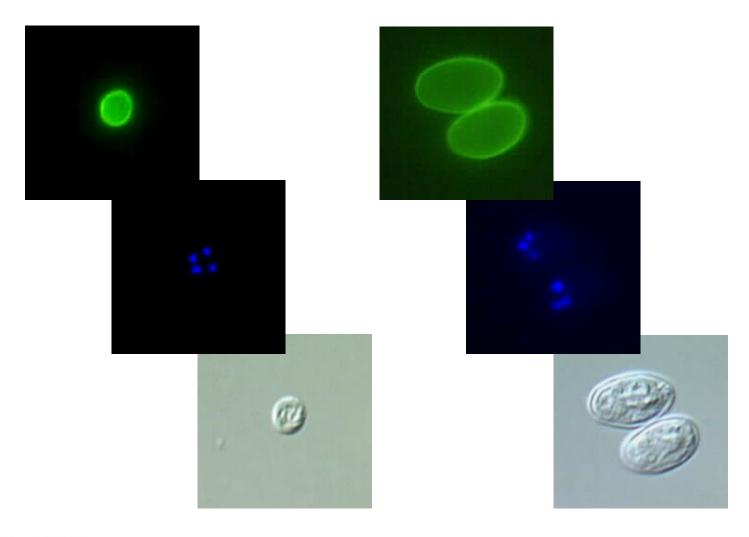


The acid is not responsible for acid dissociation!

The vortexing breaks the bond between the bead and the (oo)cyst, the acid just prevents the bond reforming!



Sample Staining



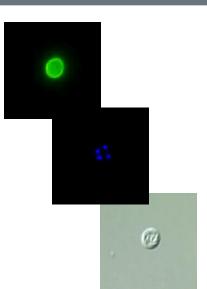


Sample Staining

- Be consistent
 - time and temperature affect staining and background
- Not all stains are created equal
 - always follow the manufacturer's instructions
- Angle slides to aid liquid removal
- Always use a pre-warmed humid chamber
- Consider using PBS rinses rather than reagent water rinses
 - particularly if it's in the instructions
- Dry slides down slowly*
 - be careful with slide warmers (some have hot-spots)
- Take care when mounting and sealing slides

^{*} personal preference is overnight at ambient temperature





Sample Staining – Liquid removal

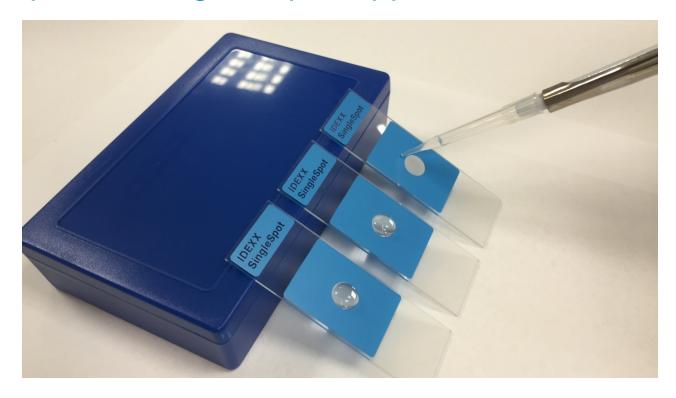




- Remove liquid as gently as possible
 - gentle aspiration
 - wicking method



Sample Staining – Liquid application



- Apply liquid as gently as possible
 - application from above the well lets gravity do the work and you don't need to change tips





Slide boxes make great humidity chambers and heat up quickly in an incubator!





A Word about Controls

- Controls are important but they don't tell you everything
- Reagent controls are important, but so are whole method controls
- Whatever you do with real samples, you should do with controls
- Spiking is key
 - if you don't introduce all of the spike dose, you're on a losing streak from the off
 - not all spikes are created equal test them out



The Key to Consistency

- If you want to have consistent results
 - YOU HAVE TO HAVE A CONSISTENT METHOD
- If you want analysts to perform consistently, give them the ability to do so:
 - have them concentrate on the right things
 - keep the controls & checks to an essential minimum
 - cut the paperwork & bureaucracy
 - don't put quantity before quality



Thank You!!!

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